# STUDIES ON THE STREPTOMYCES SC4:

# Chemical Formulation of Antibiotic SC<sub>4</sub>-X

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(Received December 3, 1982; Accepted December 29, 1982)

#### Abstract

A new species of *streptomyces*, namely *streptomyces*  $SC_4$ , was isolated from a soil sample collected in Taiwan. An antibiotic,  $SC_4$ -X, was isolated and purified as its amorphous sulfate salt from *streptomyces*  $SC_4$ . It is active against Gram-positive and Gram-negative bacteria and fungi. This antibiotic is tentatively considered to be a new member of the streptothricin type antibiotics, which contain three molecules of  $\beta$ -lysine, one molecule of aminosugar, and one molecule of streptolidine. The difference in structures between  $SC_4$ -X and those other antibiotics is the extents of aminosugar modifications. The purification and elucidation of chemical structure of this basic and water soluble antibiotic is reported.

#### Introduction

Streptothricins, first isolated by Waksman and Woodruff in 1942, are basic and water soluble antibiotics produced by Streptomyces lavendulae. Since then a number of related antibiotics have been identified, including yazumycin by Akasak et al. in 1968 and Taniyama et al. in 1971, streptothricin S15-1 by Brown et al. in 1977 and Kawamura et al. in 1976, streptolidine by Carter et al. in 1961, roseonine by Goto et al. in 1974, sclerothricin by Kono et al. in 1969, streptolin by Larson et al. in 1953 and Van Tamelen et al. in 1952, roseothricin by Nakanishi et al. in 1954, streptidine by Peck et al. in 1946, streptothricin R4H and racemomycins by Sawada et al. in 1974, glycinothricin by Sawada et al. in 1977, sclerothricin by Shimazu et al. in 1969, citromycin by Taniyama et al. in 1972, SF-701 by Tsuruoka et al. in 1968, pleocidin, mycothricin, grasseriomycin, grisin, phytobacteriomycin and polymycin by Khokhlov in 1961.

All streptothricins are active against a wide spectrum of Gram positive and Gram negative bacteria and fungi, but are highly nephrotoxic. Most streptothricins were isolated by absorption and solvent extraction. Although it has always been difficult to obtained crystalline compound in free form, crystalline derivatives have been reported as the reineckate by Fried *et al.* in 1945 and helianthate derivatives by kuehl *et al.* in 1945. Streptothricin picrate (Peck *et al.* in 1946) and sulphate (Carter *et al.* in 1954) have been also used in some isolation procedures.

Streptothricin type antibiotics are basic substances, soluble in water and dilute solutions of acid and alcohol, and decompose in concentrated acid and alkali. They are insoluble in ether, petroleum ether and chloroform. In the crude preparation, they are sensitive to heat, but withstand heating at 100°C for 15 min after purification. Treatment with proteolytic enzymes does not affect their biological activities.

Acid hydrolysis of streptothricin produced three compounds, which upon chromatography on a cellulose column with the solvent system t-butanolacetic acid-water (2:1:1, v/v) were separated and identified as L- $\beta$ -lysine (I) (Carter, Clark et al. in 1961), a cyclic guanidine derivative with the structure 2-amino-imidazoline (II), corrected by Carter and McNary as (III), and amino sugar with the structure 2-amino-2-deoxy- $\alpha$ -D-gulose (D-2-gulosamine) (IV) (Van Tameleon et al.) and its 1,6-anhydro-derivative (V) (Johnson and Westley in 1692). Complete structure of streptothricin (VI) was established by Van Tamelen et al. in 1961.

More affirmative evidence for the proposed formulae of streptothricins E-A was given by Khokhlov and Shutova. They isolated and tried to identify the products after mild hydrolysis of each streptothricin (E-A). All amide bonds in these compounds were shown to link through  $\epsilon$ -amino group of  $\beta$ -lysine residues, whereas their  $\beta$ -amino groups remain free. Uncertainty

$$NH_3$$
— $(CH_3)_3$ — $CH(NH_3)$ — $CH_3$ — $COOH$ 
(I)

COOH

$$H_2N$$
 $H_2N$ 
 $H_2N$ 
 $H_3$ 
 $H_4$ 
 $H_4$ 
 $H_5$ 
 $H_5$ 

RO ONH OH OH R, R' = H, CONH<sub>2</sub>

OR' NH 
$$n = 2,3,4,5,6$$

(COCH<sub>2</sub> CHCH<sub>2</sub> CH<sub>2</sub>CH<sub>2</sub>NH)<sub>n</sub>H

NH<sub>2</sub>

(VIIa)

remains that formulae (VII) represent the structures of all streptothricins, there is still no conclusive evidence to confirm that the location of carbamoyl group is at the C-6 carbon of the gulosamine moiety in streptothricin F(F:n=1). It is equally possible that in streptothricin E-A the carbamoyl group is located at C-3 or C-4 of gulosamine. The structures of all streptothricin E-A may therefore be designated by the general formulae (VIIa).

Streptomyces- $SC_4$  has been isolated from a soil sample collected in Kaohsiung, Taiwan. This organism is capable of producing a series of basic and water soluble antibiotics which are strongly active against the growth of Gram-positive, Gram-negative bacteria and true fungi. A new antibiotic, designated as  $SC_4$ -X, has been obtained from the fermentation broth of streptomyces- $SC_4$ . This work has been undertaken to compare the antibiotic properties and purification procedures of the aminosugar type antibiotic  $SC_4$ -X with the known streptothricin type antibiotics. The physico-chemical characteristics and chemical formulation of  $SC_4$ -X are also discussed.

#### Materials and Methods

## Strain

The strain *Streptomyces* SC<sub>4</sub> was isolated from a soil sample collected at Kaohsiung, Taiwan. According to Bergey's key, it was classified into the genus of *Streptomyces*. The organism was kept in the lyphilized form. Slants of stock cultures were maintained on trypton yeast extract glucose (TYG) agar. These slants as well as subsequent cultures used in this study were routinely incubated at 28°C for 7 days before harvest.

#### Determination of the antimicrobial spectrum

Test bacteria were cultured on the Difco antibiotics medium 1. Human pathogenic fungi were cultured on Difco Sabouraud agar. The Czapek's-Dox medium was used for plants pathogenic fungi. Antibacterial and antifungal activities of the antibiotics were measured by the paper disc method.

#### Antimicrobial assay

Bacillus subtilis PCI 219 was used as the indicator bacteria. A series of two-fold diluted antibiotic samples were prepared and each aliquot of 0.005 ml of the test solutions was applied on paper discs (0.6 cm diameter) which were then placed on agar plates seeded with appropriate organisms.

## Fermentative production

The composition of the sporulative media included 15 ml glycerol, 10 g asparagine, 5 g tyrosine, 0.5 g  $K_2HPO_4$ , 0.5 g  $MgSO_4$ , 0.01 g  $FeSO_4$ , 0.5 g NaCl and 15 g agar in 1,000 ml of deionized water. The pH was adjusted to 7.2

with 1 N NaOH before autoclaving. The composition of the seed media contained 5 g tryptone, 3 g yeast extract, 10 g glucose, 1 g K2HPO4, 1 g KH2PO4 and 15 g agar in 1,000 ml of deionized water. The fermentation media contained 15 ml glycerol, 1 g asparagine, 0.5 g tyrosine, 0.5 g K2HPO4, 0.5 g MgSO<sub>4</sub> · 7H<sub>2</sub>O<sub>5</sub> 0.01 g FeSO<sub>4</sub> · 5H<sub>2</sub>O and 0.5 g NaCl in 1,000 ml of deionized water. Each spore suspension slant was inoculated into five 250 ml Erlenmeyer flasks each containing 30 ml of the seed media. After 3 days incubation in a rotary shaker at 28°C, the inoculum (10% by volume) was transferred to 2,000 ml Erlenmeyer flasks each containing 1,000 ml seed medium. After 3 days incubation at 28°C, the inoculum (also 10% by volume) was transferred to a 15-liter jar fermentor containing 12 liters of medium. Fermentation was carried out at 28°C for 7 days under aeration of 20 liters/min and agitation at 200 rpm. During the fermentation period, 50 ml of the cultured fluid was drawn daily for the assay of antibiotic activities. The rate of growth was determined by measuring the volume of mycelia in the broth after centrifugation at 3,000 rpm for 10 minutes. The antibiotic activity of the beer supernatant was measured with Bacillus subtilis PCI 219 as the test organism.

# Isolation and purification of SC<sub>4</sub>-antibiotic

Fermentation broth (24 liters), harvested from two 15-liters fermentors, was adjusted to pH 2.0 with 6 N hydrochloric acid, stirred for 30 minutes and again adjusted to pH 6.8 with 1 N sodium hydroxide before filtration through Whatman No 1 filter paper. The filtrate was passed through Amberlite IRC 50 (H-form) column. After washing with sufficient amount of deionized water, the contents was subsequently eluted with 6 liters of 0.3 N hydrochloric acid or 10% (w/v) NaCl solutions.

The eluate was adjusted to pH 2.0 and decolorized by passing through a column containing active carbon and eluted with 70% methanol at pH 2.0. Fractions containing SC<sub>4</sub>-X were pooled and concentrated in vacuo. The concentrated solution was added with five volumes of methanol and the biologically inactive precipitates were removed through filtration. The filtrate was concentrated in vacuo and added into it twenty volumes of ehtanol-acetone mixture (1:5, v/v), yielding brown powdery precipitates containing SC<sub>4</sub>-X antibiotic. The crude antibiotic was further purified through a cellulose column (Whatman CF II) by eluting with a solvent system of *n*-propanol-pyridine-acetic acid-water (15:10:3:12, v/v). Active fractions were combined and the solvent evaporated in vacuo to dryness. Then the crude preparation was dissolved in water, passed through a column of Dowex-1 (OH-form), then adjusted the pH value of the solution to 3.0 with 2N

sulfuric acid, and finally added with twenty volumes of ethanol to precipitate  $SC_4$ -X. After filtration and drying in a desiccator, 500 mg of partially purified  $SC_4$ -Xwas obtained.

#### Purification

One gram of the partially purified SC<sub>4</sub>-antibiotics were applied on a column of Sephadex G-25 (2.5×90 cm) and eluted with deionized water. The active eluate was separated into two major fractions, designated as SC4-I and SC<sub>4</sub>-II. Pike SC<sub>4</sub>-I and SC<sub>4</sub>-II were concentrated individually in vacuo, and treated with 20 volumes of ethanol at pH 3.0 with 2N H<sub>2</sub>SO<sub>4</sub>. The later fraction, SC4-II, was chromatographed again on a carboxymethyl cellulose Whatman CM 32 column (2.5×90 cm) and eluted with a 0 to 0.4 M linear gradiant of sodium chloride. The antibiotic SC4-II was further separated into two active fractions at this step. The first fraction was designated as antibiotic SC<sub>4</sub>-X, and the second fraction was designated as antibiotic SC<sub>4</sub>-Y. Each active fraction was concentrated in vacuo and treated with charcoal for desalting. The antibiotics SC<sub>4</sub>-X and SC<sub>4</sub>-Y were eluted with 70% methanol from charcoal at pH 2.0. The purified SC<sub>4</sub>-X and SC<sub>4</sub>-Y antibiotics were confirmed as a clear single spot with ninhydrin in several silica gel TLC systems which included n-propanol-pyridine-acetic acid-water (15:10:3:12, v/v), wet-BuOH with 2% of p-toluene sulfonic acid, n-BuOH-acetic acid-water (2:1:1, v/v) and n-BuOH-pyridine-acetic acid-water-t-BuOH (15:10:3:15:4, v/v).

Proton and C-13 NMR spectra were obtained on a Joel 100 MHz spectometer. All NMR spectra were recorded at ambient temperature in  $D_2O$  and are reported as parts per million downfield from  $Me_4Si$  ( $\delta=0$ ). Routinely DSS is used as an internal standard. Infrared spectra were taken by using a Perkin-Elmer 283 B spectrophotometer as KBr pellets.

Analytical HPLC was performed by a Waters Model 6,000 A Pump equipped with Model 401 differential refractometer detector. Samples were introduced into a  $\mu$ -Bondapak-C<sub>18</sub> Column through a U-6K injector. Conditions for separation of SC<sub>4</sub>-X and A249 were specified in the results section.

Amino acid analysis was carried out by Dr. W.C. Chang, Institute of Biological Chemistry, Academia Sinica. Hydrolysate of standard protein was run for comparison.

#### Results

# General properties of SC<sub>4</sub>-X

Antibiotic SC<sub>4</sub>-X sulfate salt is a white amorphous powder which melts at 180-185°C with decomposition. It is quite stable in acidic solution but unstable in strong alkaline solution. It is hydrophilic, readily soluble in

water, slightly soluble in methanol and ethanol but insoluble in most of the common organic solvents. Ultraviolet absorption spectroscope shows no characteristic band above 220 nm. Its basic nature was indicated by paper electrophoresis. The antibiotic shows positive response to ninhydrin and Molish reaction but is negative to Benedict, Bials, Biuret, and Sakaguchi reactions.

Biological activities of SC<sub>4</sub>-X

Jan. 1983

Table 1. Antibacterial activity of SC<sub>4</sub>-X antibiotic under different pH condition

Test organisms	MIC (mcg/ml)			
rest organisms	pH 4.8	pH 7.0	pH 8.2	
Escherichia coli ATCC-10536	125	64	32	
Bacillus subtilis ATCC-6633	64	32	16	
Staphylococcus aureus ATCC-9144	125	4	1	
Sarcina lutea ATCC-9341	64	4	1	
Micrococcus flavus ATCC-10240	32	16	4	
Klebsiella pneumoniae ATCC-10031	500	16	16	
Mycobacterium pseudotuberculosis ATCC-607	64	1	1	

Paper disc method: 0.005 ml of the test solutions were applied on Whatman No 1

filter paper disc, 0.6 cm diameter.

Common bacteria: Antibiotics medium 1. 37°C for 24 hours. Mycobacterium: 5% glycerin nutrient agar, 28°C for 7 days.

Table 2. Antifungal activity of SC<sub>4</sub>-X antibiotic

Test organisms	Medium	pH 7.0	
rest organisms	Wedium		
Penicillium citrinum	C.A.	64	
Helminthosporium oryzae	C. A.	4	
Colletotrichum lagenarium	C. A.	32	
Penicillium italicum W.	C. A.	32	
Gibberella fujikuroi	C. A.	16	
Candida albicans	S. A.	16	
Cryptococcus neoformans	S. A.	32	

Paper disc method.

Method: 0.005 ml of the test solutions were applied on Watman No. 1

filter paper disc, 0.6 cm diameter.

Plant pathogenic fungi were incubated at 28°C for 48 hours.

Human pathogenic fungi were incubated at  $37^{\circ}\text{C}$  for 48 hours.

Medium: Sabouraud agar (S. A.), Czapek's-Dox agar (C. A.)

The antimicrobial activities are shown in Table 1.  $SC_4$ -X is active against Gram-positive, Gram-negative bacteria and Mycobacteria. The activitity of  $SC_4$ -X is higher in alkaline than in acidic conditions. The antifungal activities of  $SC_4$ -X are summarized in Table 2.  $SC_4$ -X is not only active against the

Table 3. The Rf values of SC<sub>4</sub>-X and streptothricin group antibiotics

C. Inc. A. C. C.	Rf values			
Solvent system	SC <sub>4</sub> -X	S15-1	A 249	SF 701
T. n-propanol-pyridine-acetic acid-water (15:10:3:12)	0.19	0.18 0.07	0.16	0.25
Ⅲ. Chloroform-methanol-28% ammonia (2:1:1)	0.44	0.42 0.36	0.35	0.63
■. n-propanol-pyridine-acetic acid-water (1:1:1:1)	0.09	0.44	0.44	0.69

Detection: Ninhydrin. Bioautography with *B. subtilis PIC 219*. TLC-plates silica gel F254 pre coated layer thickness 0.25 mm.

agricultural pathogenic fungi, Helminthosporium oryzae, and Gibberella fujikuroi but also capable of inhibiting human pathogenic fungi, Candida albicans and Cryptococcus neoformans.

# Characterization of SC<sub>4</sub>-X by TLC

The Rf values of  $SC_4$ -X and other known streptothricin-type antibiotics on silica gel TLC are shown in Table 3. The Rf value of  $SC_4$ -X is different from other known streptothricin antibiotics. The Rf value of  $SC_4$ -X on silica gel TLC developed with the solvent of n-propanol-pyridine-acetic acidwater (1:1:11, v/v) is only 0.1 which is much smaller than those of streptothricin antibiotics S15-1, A249 and Sf 701.

# Chromatographic characterization of SC<sub>4</sub>-X by HPLC

A comparison of purity and identities of  $SC_4$ -X with other known streptothricin type antibiotics by HPLC is shown in Fig. 1. A249 and S15-1 each contains at least three peaks. In  $SC_4$ -X, the predominant component is peak A which has similar Rf value with a minor component in S15-1. Small amount of highly purified  $SC_4$ -X was obtained by repeated injection and collection of the predominant peak in  $SC_4$ . The purity of  $SC_4$ -X thus collected was shown in Fig. 1, which indicates that the impurity peak is trivial. The negative peak in this chromatogram is due to solvent.

Hydrolysis of SC<sub>4</sub>-X and S15-1 by 6 N HCl in sealed bomb tubes at 110°C were carried out at various time intervals. Result from amino acid analysis

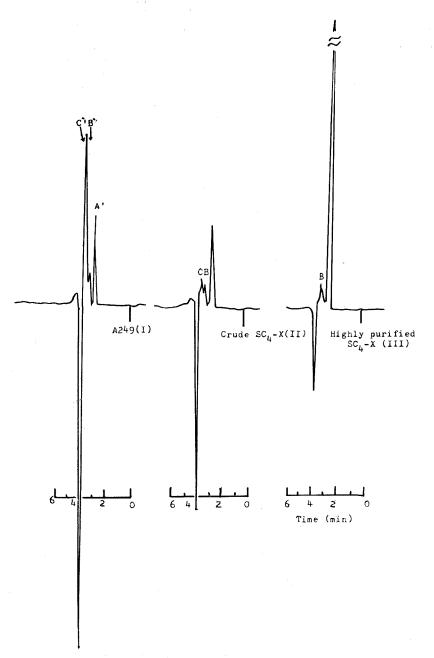


Fig. 1. Chromatogram of A249 (I), Crude  $SC_4$ -X (II), and highly purified  $SC_4$ -X (III). Separation was carried out by a  $\mu$ -Bondapak C-18 column (30×0.4 cm) solvent: 1% HOAc. Peaks were detected by a refractometer, flow rate: 1 ml/min

of  $SC_4$ -X hydrolysate is shown in Fig. 2. It evidently shows the absence of any other amino acid except two peaks. The first peaks moves with Rf value very similar to histidine and the second one has almost identical Rf value with  $\beta$ -lysine. The peak comes out last from the column is found to be ammonia. The ratio of the peak areas of these three components is approximately 1:3:1 in forty-eight hour hydrolysate. Result of the hydrolysate by HPLC is reported in Fig. 3.

# Spectroscopic characterization

Since  $SC_4$ -X sulfate is very hydroscopic, infrared spectrum of  $SC_4$ -X was taken in KBr pellet immediately after prolonged drying in vacuo (Fig. 4). It shows the presence of -NH, C-OH, and amides which are also present in the IR spectrum of S15-1 (Fig. 5).

Chemical shift and tentative assignment of C-13 NMR spectra of SC<sub>4</sub>-X

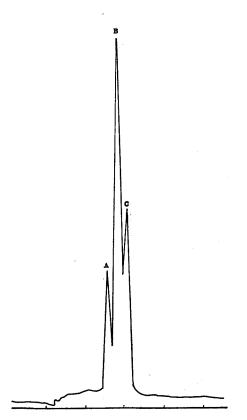


Fig. 2. Chromatogram of the hydrolysate of  $SC_4$ -X antibiotic by using an amino acid analyzer.

Peak A: Streptolidine. Peak B: β-lysine. Peak C: Ammonia.

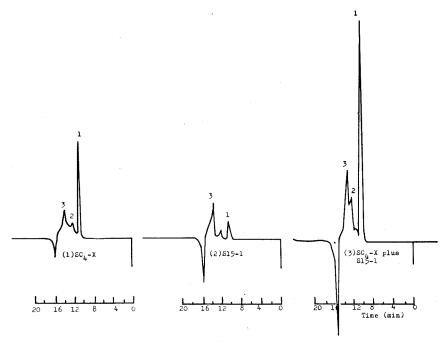


Fig. 3. HPLC analysis of the hydrolysates of  $SC_4$ -X, S15-1 and  $SC_4$ -X plus S15-1. Hydrolysates of 0.5 mg antibiotic with 6N HCl at 110°C for 24 hours in a sealed

Detector:  $\Delta$  IR × 32, Solvent: 1% HOAc/H<sub>2</sub>O, Column:  $\mu$ -Bondapak C<sub>18</sub>,

Flow rate: 1 ml/min, Chart speed: 0.1 in/min.

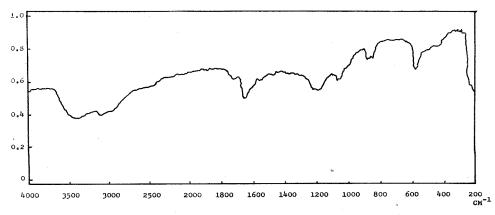
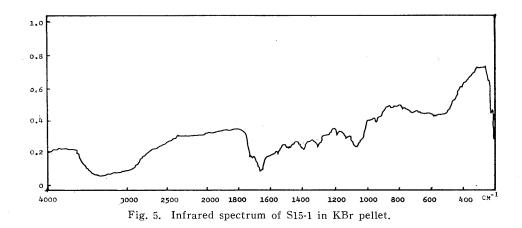


Fig. 4. Infrared spectrum of SC<sub>4</sub>-X in KBr pellet.



**Table 4.** Chemical shifts and tentative assignment of C-13 magnetic resonances of SC<sub>4</sub>-X, A249 and S15-1

	SC <sub>4</sub> -X	A 249	S 15-
	49.4	49.2	49.4
	37.4	37.1	37.4
β-lysine	29.9	29.9	29.9
	23.8	23.8	23.8
	39.8	39.8	39.9
	55.2	55.2	55.2
	61.7	61.7	61.7
Sugar	68.6	67.6	67.6
ougai	70.8	70.8	70.8
	74.3	74.3	74.3
	79.6	79.5	79.6
	49.8	50.1	49.8
Streptolidine	50.1	50.3	50.1
	61.2	61.1	61.2
·	61.5	61.5	61.5
	158.3	158.4	158.8
Carbamoyl and Quanidino	_	163.2	163.7
C=0	172.2	172.6	172.8
	172.4		
	172.8		

- 1) Chemical shifts are reported in ppm,  $\delta$  (Dioxane=67.4)
- 2) The resonances of streptolidine D-glucosamine, carbamoyl, guanidino and carbomyls have not yet been satisfactorily assigned.
- 3) 100 MHz Joel FT-NMR Spectrometer.

and other related streptothricins are shown in Table 4. The presence of peaks corresponding to  $\beta$ -lysine is particularly grouped at the upper portion. Values of the chemical shift of  $\beta$ -lysine and streptolidine portion were found identical with other streptothricin antibiotics such as S15-1 and A249.

#### Discussion

 $SC_4$ -X antibiotic is a water-soluble basic antibiotic. As shown in Table 5, it is different from known antibiotics.  $SC_4$ -X inhibits not only *Mycobacterium*, Gram-positive and Gram-negative bacteria but also human and plants pathogenic fungi. It is therefore not identical with streptomycin, viomycin, neomycin and kanamycin. The chemical tests, Rf value of TLC or HPLC chromatography, and antimicrobial spectrum of  $SC_4$ -X suggested us that it is probably a new member of streptothricin-like antibiotics.

Streptomyces  $SC_4$  produces a series of compounds with broad spectrum of antibiotic activities. We have concentrated in this report on the structural elucidation of  $SC_4$ -X.

Result from the amino acid analysis of  $SC_4$ -X hydrolysate indicates the presence of compound A, lysine analog (B), and ammonia. The compound (B) is eventually confirmed by NMR to be  $\beta$ -lysine. The compound A, which behaves like histidine in amino acid analysis, is actually streptolidine. In the forty-eight hours hydrolysate of  $SC_4$ -X by 6 N HCl. The product ratio of  $\beta$ -lysine: Streptolidine: NH<sub>3</sub> is approximately 3:1:1. In conjunction with other chemical tests,  $SC_4$ -X is evidently a streptothricin type antibiotic. Interpretation of proton and C-13 NMR spectra further support that the lysine moieties are  $\beta$ -lysine and an intact streptolidine is present. The C-13

Antibiotics		Color reaction			Dr
	Maltol	Ninhydrin	Sakaguchi	m.p.	Rf values*
SC <sub>4</sub> -X	_	+	_	180-185	0.44
Yazumycin	_	+	+	230-236	0.3-0.33
Streptothricin	+	+		213-217	0.26
Zygomycin A					0.62
Viomycin	-				0.11
Paromomycin					0.68
Aminosidin					0.68
Glebomycin		_			

Table 5. Comparison of SC<sub>4</sub>-X with other known antibiotics

Solvent: the upper layer of CHCl<sub>3</sub>:MeOH:17% NH<sub>4</sub>OH (2:1:1, v/v).

<sup>\*</sup> Rf value by silica gel T.L.C.

NMR assignment of  $\beta$ -lysine and streptolidine is reported.

The presence of a highly modified six-membered ring sugar moiety is also confirmed by NMR and chemical tests. Difference in 50-75 ppm region of C-13 NMR exists particularly between S15-1 and SC<sub>4</sub>-X implies that the configuration and/or modification, including linkage of  $\beta$ -lysine and carbamyl moieties, could be different.

Streptothricin type antibiotics are extremely difficult to purify to homogeneity. Traditional separation based on size and charge is hard to distinguish cluster of compounds with structural difference merely on the number of  $\beta$ -lysine molecules and positions of sugar modification. In our comparison study for purity by HPLC of some characterized streptothricin type antibiotics, namely A249 and S15-1, we found that A249 and S15-1 each contains at least three compounds. The HPLC profiles of S15-1 and SC<sub>4</sub>-X are almost identical except the peaks corresponding to  $\beta$ -lysine reflect the numbers of  $\beta$ -lysine moieties of individual antibiotics.

In general, we have confirmed that  $SC_4$ -X is a streptothricin type antibiotics which is comprised of one modified sugar, one streptolidine and 3 molecules of  $\beta$ -lysine. Our results is not sufficient to clearly assign the stereochemistry and linkage of the subtituents of pyranose moiety. Further improvement of purification techniques for larger scale sample is definitely a obligatory step to secure complete assignment of the fine structure of  $SC_4$ -X.

# Acknowledgement

The authors wish to express their sincere thank to Dr. W.E. Brown, Squibb Institute of Medical Research, Princeton, New Jersey, U.S.A. for his generous supply of streptothricin S15-1 (SQ21, 704) and to Dr. T. Tsuruoka, Research Laboratories, Meiji Seika Kaisha Ltd, Kohoku-Ku, Yokohama, Japan for antibiotics SF701, A249 and  $\beta$ -lysine.

This work was supported in Part by a grant from the National Science Council and Chung-Cheng Funds, Republic of China in 1982.

Paper No. 258 of the Scientific Journal series, Institute of Botany, Academia Sinica.

#### References

Akasak, K., H. Abe, A. Seino and S. Shirato. 1968. Yazumycin, a new antibiotic produced by *Streptomyces lavendulae*. J. of Antibiotics, 21: 98-105.

Aszalos, A., S. Davis and D. Frost. 1968. Classification of crude antibiotics by instant thin-layer chromagraphy (ITLC). J. of Chromatography, 37: 487-498.

Brown, W. E., J. Szanto, E. Meyers, T. Kawamura and K. Arima. 1977. Taeniacidel activity of Streptothricin antibiotic complex S15-1 (SQ21, 704). J of Antibiotics, 30: 886-889.

- Carter, H.E., R.K. Clark, P. Koine, J.W. Rothrock, W.R. Tayler, C.A. West, G.B. Whitfield and G. Jackson. 1954. Streptothricin I. preparation, properties and hydrolysis products. J. Am. Chem. Soc., 76: 566-569.
- Carter, H. H., C. C. Sweeley, E. E. Daniels, J. E. Mcnary, C. P. Schaffner, C. A. West, E. E. Tamelen, J. R. Dyer and H. A. Whaley. 1961. Streptothricin and streptolin: The structure of sterptolidine (Roseonine). J. Am. Chem. Soc., 83: 4296-4297.
- Fried, J. and O. Wintersteiner. 1945. Crystalline reineckates of streptothricin and streptomycin. Science, 101: 613-615.
- Goto, T. and T. Ohgi. 1974. Synthesis of roseonine (streptolidine), a guanidino-amino acid component of streptothricin group antibiotics. Tetrahedran Letters, 15: 1413-1416.
- Horowitz, H. and C.P. Schaffner. 1958. Paper chromatography of streptothricin antibiotics. Anal. Chem., 30: 1616-1620.
- Ito, Y., Y. Ohashi, Y. Sakurai, M. Sakurazawa, H. Yoshida, S. Awataguchi and T. Okuda. 1968. New basic water-soluble antibiotics BD-12 and By-81 II Isolation, purification and properties. J. of Antibiotics, 21: 307-313.
- Johnson, A.W. and J.W. Westley. 1962. Streptothricin group of antibiotics. Part 1. The general structural pattern. J. Chem. Soc., 1642-1652.
- Jeanloz, R.W. 1959. The synthesis of 2-amino-1, 6-anhydro-2-deoxy-β-gulopyranose hydrochloride. J. Am. Chem. Soc., 81: 1956-1960.
- Kawamura, T., K. Tago, T. Beppu and K. Arima. 1976. Antiviral antibiotic S15-1. J. of Antibiotics, 29: 242-247.
- Kawamura, T., T. Kimura, T. Tago, T. Geppn and K. Arima. 1976. The identity of S15-1-A and B with racemomycins A and C. J. of Antibiotics, 29: 845-846.
- Khokhlov, A.S. and P.D. Reshetov. 1964. Chromatography of streptothricins of carboxy-methylcellulose. J. Chromatography, 14: 495-496.
- Khokhlov, A.S. and K.I. Shutova. 1972. Chemical structure of streptothricins. J. of Antibiotics, 25: 501-508.
- Khokhlov, A.S. 1961. Chemistry of antibiotics. Publ House of USSR, Acade. Sci. Moscow. Komatsu, N., Y. Saburi, Y. Hirata, T. Mizuno, and N. Sakai. 1952. Countercurrent distribution studies of H-277. J. of Antibiotics, 9: 522-523.
- Kono, Y., S. Makino, S. Takenchi and H. Yonehara. 1969. Sclerothricin, a new basic antibiotic. J. of Antibiotics, 22: 583-589.
- Kuehl, F. A., R. L. Peck, A. Walti, K. Folkers. 1945. Streptomyces antibiotics. I. Crystalline salts of streptomycin and streptothricin. Science, 102: 34-35.
- Larson, L. M., H. Sternberg and W. H. Peterson. 1953. Production, isolation and components of the antibiotic streptolin. J. of Am. Chem. Soc., 75: 2036-2039.
- Nakanishi, K., T. Ito and Y. Hirata. 1954. Structure of a new amino acid obtained from roseothricin. J. Am. Chem. Soc., 76: 2845-2846.
- Peck, R. L., A. Walti, R. P. Graber, E. Flynn, C. E. Hoffhine, V. Allfrey and K. Folkers. 1946. *Streptomyces* antibiotics. VI. Isolation of streptothricin. J. Am. Chem. Soc. 68: 772-776.
- Peck, R. L., A. Walti, R. P. Graber, E. Flynn, C. E. Hoffhine, V. Allfrey and K. Folkers. 1946. Streptomyces antibiotics VII. The structure of streptidine. J. Am. Chem. Soc., 68: 776-781.
- Peterson, D. H., D. R. Colingsworth, L. M. Reineke and C. Deboer. 1947. Evidence for the presence of streptothricin in streptolin culture filtrates. J. Am. Chem. Soc., 69: 3145-3146.
- Peterson, D.H., and L.M. Reineke. 1950. A paper chromatographic technique and its application to the study of new antibiotics. J. of Am. Chem. Soc., 72: 3598-3603.
- Peterson, D.H. and H.A. Sober. 1956. Chromatography of proteins. I. Cellulose ion-exchange adsorbents. J. Am. Chem. Soc., 78: 751-755.
- Rivett, R.W. and D.H. Peterson. 1947. Streptolin, a new antibiotic from a species of streptomyces. J. Am. Chem. Soc., 69: 3006-3009.

- Sawada, Y., H. Taniyama, N. Hanyuda, H. Hayashi and T. Ishida. 1974. A new strepto-thricin antibiotic R 4H. J. of Antibiotics, 27: 535-543.
- Sawada, Y., H. Sakamoto and H. Taniyama. 1974. Studies on chemical modification of streptothricin-group antibiotics III. Partial N-acetylation of racemomycin and their biological activity. J. of Pharmaceutical Society of Japan, 94: 176-180.
- Sawada, Y. and H. Taniyama. 1974. Studies on chemical modification of streptothricingroup antibiotics. IV. Preparation of  $\beta$ -N-Acetylracemomycin-A derivative and its antimicrobial activity. J. of Pharmaceutical Soc. of Japan, 94: 264-266.
- Sawada, Y. and H. Taniyama. 1974. Studies on chemical modification of streptothricingroup antibiotics. V. Synthesis of amino acid derivative on β-lysine of racemomycin A and their biological activity. J. of the Pharm. Soc. of Japan, 94: 858-864.
- Sawada, Y., S. Kawakami and H. Taniyama. 1977. Glycinothricin, a new streptothricinclass antibiotics from *Streptomyces griscus*. J. of Antibiotics, 30: 460-467.
- Shimazu, A., T. Hidaka, S. Otsuka, M. Nishiyama and H. Yonehara. 1969. Streptomyces sclerogranulatus sp. Nov., The producer of sclerothricin. J. of Antibiotics, 22: 590-596.
- Shoji, S., S. Kozuki, M. Ebata and H. Otsuka. 1968. A water-soluble basic antibiotic E-749-C identical with LL-AC 541. J. of Antibiotics, 21: 509-511.
- Swart, E.A. 1949. The use of counter-current distribution for the characterization of *Streptomyces* antibiotics. J. Am. Chem. Soc., 71: 2942-2944.
- Tamelen, E.E. and E.E. Smissman. 1952. Streptolin, the structure and synthesis of isolysine. J. Am. Chem. Soc., 74: 3713-3714.
- Tamelen, E.E. and E.E. Smissman. 1953. Streptolin, the structure and synthesis of isolysine. J. Am. Chem. Soc., 75: 2031-2035.
- Tamelen, E. E., J. R. Dyer, H. E. Carter, J. V. Pierce and E. E. Daniels. 1956. Structure of the aminosugar derived from streptothricin and streptolin B. J. Am. Chem. Soc., 78: 4817-4818.
- Van Tamelen, E. E., S. R. Dyer, H. A. Whaley, H. E. Carter and G. B. Whitfield. 1961. Constitution of the streptolin-streptothricin group of *Streptomyces* antibiotics. J. Am. Chem. Soc., 83: 4295-4296.
- Taniyama, H. and S. Takemura. 1957. Chemical studies on antibiotics produced by *Actinomycetes*. I. Racemomycin (1), Isolation and purification of racemomycin B (229-B). J. of Pharm. Soc. of Japan. 77: 1210-1214.
- Taniyama, H. and S. Takemura. 1957. Chemical studies on antibiotics produced by Actinomycetes. II. Racemomycin. (2). Hydrolysis of racemomycin B. (i). J. of Pharm. Soc. of Japan. 77: 1215-1217.
- Taniyama, H. and S. Takemura. 1958. Chemical studies on antibiotics produced by *Actinomycetes*. III. Racemomycin (3). Hydrolysis of racemomycin B (ii). J. of Pharm. Soc. of Japan, 78: 742-744.
- Taniyama, H., Y. Sawada and T. Kitagawa. 1971. Chromatography of racemomycin on dextran gel. J. of Chromatogr. 56: 360-362.
- Taniyama, H., Y. Sawada and LT. Kitagawa. 1971. Studies on the inactivation and regeneration of streptothricin. J. of Antibiotics, 24: 662-666.
- Taniyama, H., Y. Sawada and T. Kitagawa. 1971. The identity of yazumycins A and C with racemomycins A and C. J. of Antibiotics, 24: 390-392.
- Taniyama, H., Y. Sawada and K. Hasimoto. 1972. Studies on chemical modification of streptothricin-group antibiotics. I. On citromycin derivatives. J. of the Phar. Soc. of Japan, 92: 182-186.
- Tarasiejska, Z. and R. W. Jeanloz. 1957. The synthesis of D-gulosamine. J. Chem. Soc., 79: 2660-2661.
- Tarasiejska, Z. and R.W. Jeanloz. 1957. The synthesis of D-gulosamine hydrochloride. J. Am. Chem. Soc., 79: 4215-4218.
- Tsuruoka, T., T. Shoumura, N. Ezaki, T. Niwa and T. Nhda. 1968. SF-701, A new streptothricin-like antibiotic. J. of Antibiotics, 21: 237-238.

- Vander Brook, M. J., A. N. Wick, W. H. Devries, R. Harris and G. F. Cartland. 1946. Extraction and purification of streptomycin, with a note on streptothricin. J. of Biological Chemistry, 165: 463-468.
- Waksman, S. A. and H. B. Woodruff. 1942. Streptothricin a new selective bacteriostatic and bactericidal agent, particularly active against gram-negative bacteria. Proceeding of the Society for Experimental Biology and Medicin., 49: 207-210.
- Waksman, S. A. 1943. Production and activity of streptothricin. J. of Bacteriology, 46: 299-310.

# 鏈 黴 菌 SC<sub>4</sub> 之 研 究SC<sub>4</sub>-X 抗生素之化學組成

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Streptomyces sp. (簡稱爲鏈黴菌  $SC_4$ ) 爲由臺灣土壤中分離而得之新抗生素產生菌。其所產生的抗生素抗菌譜甚廣,對革蘭氏陽性、陰性菌及動植物病原性黴菌等具有生長抑制之作用。 $SC_4$ -X 抗生素可將其培養液,經離子交換及層柱分析法予以純化。經紅外光譜,氫及碳13核磁共振譜,高效液相及薄層層析法及胺基酸分析法之結果得知  $SC_4$ -X 之化學結構與 streptothricin 抗生素之 A249 及 S15-1 類似但不儘相同。 $SC_4$ -X 之分子內含有 3 分子之 $\beta$ -lysine,1 分子之胺醣及 1 分子之 streptolidine 但可能在胺醣化學修飾及立體化學上略有不同。本文即探討  $SC_4$ -X 之生物活性及其化學結構。