

# *Pileomyces formosanus* gen. et sp. nov. (Halosphaeriaceae, Ascomycota) from a rocky shore of Taiwan

Ka-Lai PANG\* and Jen-Sheng JHENG

*Institute of Marine Biology, National Taiwan Ocean University and Center of Excellence for Marine Bioenvironment and Biotechnology, 2 Pei-Ning Road, Keelung 20224, Taiwan*

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**ABSTRACT.** In Taiwan, knowledge of marine fungi is fragmentary with recent studies documenting 69 species from mangrove habitats, of which 34 taxa were new records for the country. Marine fungi on trapped substrates from rocky shores of Taiwan are comparatively less known. In a fieldtrip to Yingkeshih (New Taipei City, northeastern Taiwan), a new fungus belonging to the Halosphaeriaceae was discovered. This fungus is characterized by dark-coloured ascomata, broadly clavate asci with an apical thickening, pore and retraction of plasmalemma, and ellipsoidal ascospores with a sheet-like appendage covering one end, which eventually detaches from the ascospore. This distinctive ascospore appendage morphology is unique in the Halosphaeriaceae and *Pileomyces formosanus* gen. et sp. nov. is introduced to accommodate this fungus. *Pileomyces* resembles *Aniptodera* and *Phaeonectriella* in the morphology of the ascus, and *Tirisporella* (*T. beccariana*) in gross morphology of the ascospore appendage, however, the appendage is attached to the ascospores at the apex in *Tirisporella*.

**Keywords:** Aquatic fungi; Ascomycota; Microascales; Taxonomy.

## INTRODUCTION

Obligate marine fungi grow and sporulate exclusively in a marine or marine-related habitat (Kohlmeyer and Kohlmeyer, 1979). Habitats of marine fungi include rocky shores, sandy beaches, mangrove forests, and marine fungi occur primarily on woody tissues, but leaves, fruits, reeds, sea grasses, algae and seaweeds are also substrates (Vrijmoed, 2000). Marine fungi can also be isolated from animal exoskeletons, keratinaceous substrates, sediments, sea foam and seawater (Vrijmoed, 2000). Marine fungi play a major role in the decomposition of complex organic matter in the marine ecosystem by producing wood-modifying enzymes (cellulases, laccases and Mn-dependent peroxidases) that degrade lignocellulosic materials (Bucher et al., 2004; Luo et al., 2005). All contribute to the production of particulate and dissolved organic matter for the food web of the oceans (Jones and Pang, 2010).

Diversity of marine mangrove fungi is comparatively well-known in Taiwan (Pang et al., 2010; 2011b), however, information on their diversity on rocky shores is scarce. Hsieh et al. (2002) documented marine fungi from rocky shores of Taiwan, many of which were typical temperate water species, including *Halosphaeria appendiculata* and *Remispora* spp. (Koch and Petersen, 1996). During a field-

trip to a rocky shore in northeastern Taiwan, a fungus with characteristics of the Halosphaeriaceae was discovered. Ascospore appendage of this taxon was distinctive and significantly different from other genera in the family, and this therefore warrants the establishment of a new genus.

## MATERIALS AND METHODS

Drift and trapped bamboo/wood was collected at Yingkeshih, New Taipei City, Taiwan, on 6 September 2010. Samples were transported to the laboratory and incubated on a tray lined with moist tissue paper for up to 1 month. Fruiting bodies were observed using an Olympus SZ61 stereomicroscope (Tokyo, Japan), sectioned with a razor blade, and centrum material mounted on a slide in sterile seawater. Morphology of the asci and ascospores was observed using an Olympus BX51 microscope (Tokyo, Japan). Photographs were taken with an Olympus DP20 Microscope Camera (Tokyo, Japan).

Small pieces of decaying bamboo with ascomata of the new fungus were fixed by immersion in FAA (50% ethanol, 5% glacial acetic acid and 5% paraformaldehyde) overnight at 4°C. Fixed samples were washed three times with 50% ethanol. Samples were then dehydrated in a graduated ethanol and 1-butanol series, infiltrated gradually and embedded in paraffin. Paraffin sections (8-10 µm) were cut on a FRM-200P rotary microtome (Japan), floated over a water-bath at 42°C to relax sections, and mounted on slides. Dried sections were deparaffinised and

\*Corresponding author: E-mail: klpang@ntou.edu.tw; Tel: +886-02-24622192#5319; Fax: +886-02-24633152.

rehydrated through a graded series of ethanol. Sections were stained with 1% safranin O in 50% ethanol (10 sec) and 0.5% orange G in 95% ethanol (30 sec). After washing and dehydration, each stained section was permanently mounted with a cover slip and Permount (Fisher, Fair Lawn, USA). Specimens were observed under the microscope and photographed.

## Taxonomy

***Pileomyces*** K.L. Pang & Jheng, gen. nov. (Mycobank MB519625)

Ascomata fusca, pyriforma, immersa, erumpentia vel exposita, coriacea, ostiolata, laevia. Ostiola brevicollia. Periphyses absentes. Peridium fuscum, unistratum, constans ex cellulis elongatis stratorum plurium. Asci unitunicati, late clavati, tunica sub apice incrassati, apice poro penetrati, coma brevi, leptodermi, octospori, persistentes vel semi-persistentes, ex pulvino cellularum pseudoparenchymatarum ad basim ascomati orientes. Catenophyses praesentes, irregulares. Ascospores ellipsoideae cum apice rotundus, uniseptatae, hyalinae, laeves, leptodermiae. Appendices praesentes, ad unum polum.

***Typus generis: Pileomyces formosanus*** K.L. Pang & Jheng

Ascomata dark-coloured, pyriform with globose to subglobose venter, immersed, erumpent or exposed, coriaceous, ostiolate, smooth. Ostioles short-necked. Periphyses absent. Peridium dark-coloured, one-layered, composed of a few layers of elongated cells. Asci unitunicate, broadly clavate, apically thickened with retraction of plasmalemma, with an apical pore, short pedunculate, thin-walled, eight-spored, persistent or semi-persistent, developing from inner wall of ascomal base. Catenophyses present, irregular in shape. Ascospores ellipsoidal with rounded apices, one-septate, hyaline, smooth, thin-walled. Appendages initially covering one end of ascospores, gradually detached from the ascospore wall to form an ellipsoidal sheet.

*Etymology:* From the Latin 'pileus', in reference to the shape of the ascospore appendage.

*Anamorph:* Unknown.

***Pileomyces formosanus*** K.L. Pang & Jheng, sp. nov. (Mycobank MB563712). Figures 1-8

Ascomata 221-(231)-241 × 182-(234)-285 μm (n=2), fusca, pyriforma, immersa, erumpentia vel exposita, coriacea, ostiolata, laevia. Ostiola brevicollia. Periphyses absentes. Peridium 15-(19)-24 μm (n=4), fusca, unistratum, constans ex cellulis elongatis stratorum plurium. Asci 98-(111)-128 × 27-(31)-37 μm (n=15), unitunicati, late clavati, tunica sub apice incrassati, apice poro penetrati, coma brevi, leptodermi, octospori, persistentes vel semi-persistentes, ex pulvino cellularum pseudoparenchymatarum ad basim ascomati orientes. Catenophyses

praesentes, irregulares. Ascospores 26-(30)-36 × 9-(10)-12 μm (n=50), ellipsoideae cum apice rotundus, uniseptatae, hyalinae, laeves, leptodermiae. Appendices praesentes, ad unum polum.

Ascomata 221-(231)-241 × 182-(234)-285 μm (n=2), dark-coloured, pyriform with globose to subglobose venter, immersed, erumpent or exposed, coriaceous, ostiolate, smooth. Ostioles short-necked. Periphyses absent. Peridium 15-(19)-24 μm (n=4), dark-coloured, one-layered, composed of a few layers of elongated cells. Asci 98-(111)-128 × 27-(31)-37 μm (n=15), unitunicate, broadly clavate, apically thickened with retraction of plasmalemma, with an apical pore, short pedunculate, thin-walled, eight-spored, persistent or semi-persistent, developing from inner wall of ascomal base. Catenophyses present, irregular in shape. Ascospores 26-(30)-36 × 9-(10)-12 μm (n=50), ellipsoidal with rounded apices, one-septate, hyaline, smooth, thin-walled. Appendages initially covering one end of ascospores, gradually detach from ascospore to form an ellipsoidal sheet.

*Etymology:* In reference to Taiwan where the holotype was collected.

*Anamorph:* Unknown.

*Holotype:* TAIWAN: Yingkeshih. On a piece of unidentified trapped bamboo culm, 6 September 2010, J.S. Jheng & K.L. Pang, BBH 30192 (BIOTEC Bangkok Herbarium), dried bamboo.

*Sequence:* JX003862 (internal transcribed spacers (ITS) region of the nuclear rDNA, NCBI).

*Known geographical distribution:* Yingkeshih, Taiwan.

*Substrata:* Trapped bamboo on rocky shore.

## RESULTS AND DISCUSSION

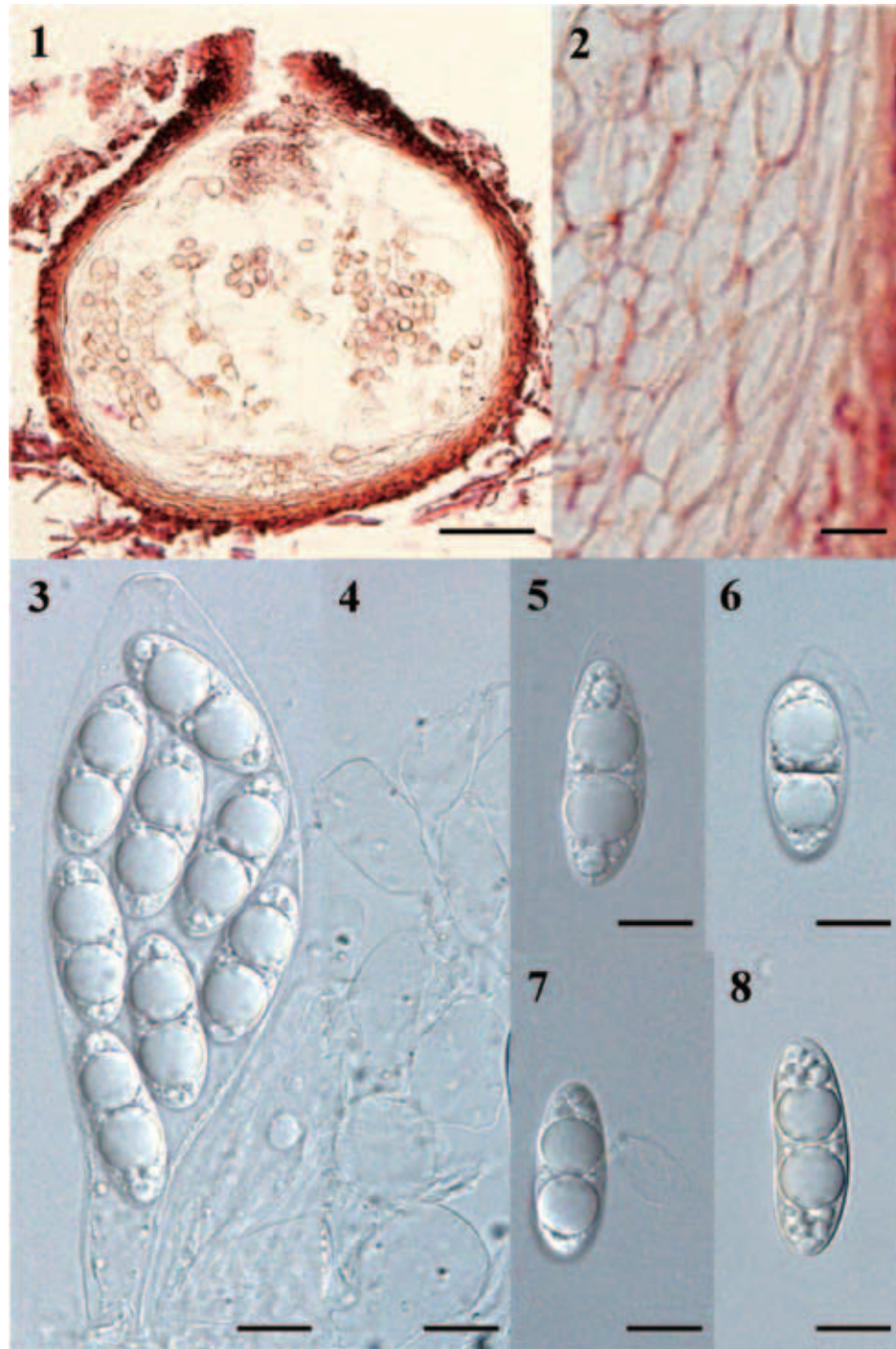
Jones et al. (2009) recorded 530 marine fungi with most species described in the period 1980-1989 and 1990-1999 (291 species), but in recent years, this dramatically decreased. Jones (2011) recently estimated the total number of marine fungi may be in excess of 10,000 and suggested the exploration of wider geographical locations for marine fungi. Pang et al. (2011a) recently recorded six lignicolous marine fungi from an environmentally extreme location, Longyearbyen, inside the Arctic circle and four of these were new to science (Pang et al., 2008; 2009; 2011a).

Taiwan has a long coastline, making it an ideal place for the study of marine fungi. The western Taiwan coast is dominated by mangrove habitats while the eastern and northeastern coasts consist of rocky shores. Pang et al. (2011b) documented 69 species of marine fungi from mangrove habitats of Taiwan while Hsieh et al. (2002) recorded 26 taxa on driftwood from Taiwan rocky shores, making an inventory of 95 taxa. However, many new taxa have been discovered in Taiwan for the last few years and await formal description (K.L. Pang, unpublished results).

The new fungus is a typical member of the Halosphaeriaceae, with its perithecioid ascomata, pseudoparen-

chymatous centrum, broadly clavate asci, presence of catenophyses, and ellipsoidal/fusiform, mostly hyaline ascospores with appendages (Spatafora et al., 1998). Ascus of *P. formosanus* shows great resemblance with that of *Aniptodera* and *Phaeonectriella*. In *Aniptodera*, ascospores are generally thick-walled, with/without unfurling as-

cospore appendages (Shearer and Miller, 1977; Hyde et al., 1999), while ascospores of *Phaeonectriella* are dark-coloured, with/without unfurling ascospore appendages (Eaton and Jones, 1971; Hyde et al., 1999). The distinctive character of *P. formosanus* is the sheet-like appendage on one end of the ascospores. However, mechanism on the



**Figures 1-8.** *Pileomyces formosanus* gen. et sp. nov. (holotype). 1, Pyriform ascoma with a subglobose venter; 2, One-layered peridium of elongate cells; 3, Clavate ascus with an apical thickening, a pore and retraction of plasmalemma; 4, Catenophyses; 5, Ascospore with sheet-like appendage on one end; 6, Appendage starting to detach from ascospore when mounted in seawater; 7, Appendage entirely detached from ascospore forming an ellipsoidal sheet; 8, Ascospore without an appendage. Scale bars: 1= 50  $\mu$ m, 2=5  $\mu$ m, 3-8=10  $\mu$ m.

formation of this ascospore appendage is unknown. The ascospore appendage was not visible inside the ascus, but was evident covering one end of the ascospores (Figure 5) when they were mounted in seawater. The appendage is not attached to the ascospore by a structure, and detaches from it to form an ellipsoidal sheet. This appendage does not appear to be gelatinous in nature and does not dissolve when mounted in seawater. An ultrastructural study of ascospores of *P. formosanus* is required to detail formation of the appendage. *Corollospora maritima* and *Chaetosphaeria chaetosa* also have sheet-like ascospore appendages resulting from fragmentation of a sheath, but are still attached to the spore at the equatorial position (Jones and Moss, 1978; Jones et al., 1983). In *Tirisporella beccariana*, the sheath-like apical appendage at one end of the ascospores is secreted through mesosporium (Jones et al., 1996). Gross morphology of ascospore appendage of *P. formosanus* and *T. beccariana* is extremely similar, however, the appendage is attached to the ascospores at the apex in the latter.

Many genera in the Halosphaeriaceae have ascospores with unipolar appendages including *Moana* Kohlm. & Volkm.-Kohlm., *Oceanitis* Kohlm. (*O. cincinnatula* (Shearer & J.L. Crane) J. Dupont & E.B.G. Jones, *O. unicaudata* (E.B.G. Jones & Le Camp.-Als.) J. Dupont & E.B.G. Jones), *Okeanomyces* K.L. Pang & E.B.G. Jones, *Ophiodeira* Kohlm. & Volkm.-Kohlm. and *Tirisporella* E.B.G. Jones & Vrijmoed. However in these genera (with the exception of *Okeanomyces*), the appendages are hamate, which gradually unfurl to form long thread-like, sticky appendages and therefore differ from that found in *Pileomyces*. The ascospore appendage of *Pileomyces* also resembles that of *Okeanomyces*. *Okeanomyces* (*O. cucullatus*) is characterized by brownish-black to brownish-red ascomata, deliquescent asci and cylindric-fusiform ascospores with a single deciduous, cap-like or globose gelatinous appendage (Kohlmeyer, 1964). *Pileomyces* differs from *Okeanomyces* in its persistent/semi-persistent asci with an apical thickening, a pore and retraction of plasma-lemma and sheet-like appendage which detaches from the ascospores to form an ellipsoidal sheet.

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## LITERATURE CITED

- Bucher, V.V.C., K.D. Hyde, S.B. Pointing, and C.A. Reddy. 2004. Production of wood decay enzymes, mass loss and lignin solubilisation in wood by marine ascomycetes and their anamorphs. *Fungal Divers* **15**: 1-14.
- Eaton, R.A. and E.B.G. Jones. 1971. New fungi on timber from water-cooling towers. *Nova. Hedw.* **19**: 779-788.
- Hsieh, S.Y., G.F. Yuan, and H.S. Chang. 2002. Higher Marine Fungi from Taiwan. Food Industry Research & Development Institute, Hsinchu, Taiwan, R.O.C.
- Hyde, K.D., W.H. Ho, and C.K.M. Tsui. 1999. The genera *Aniptodera*, *Halosarpheia*, *Nais* and *Phaeonectriella* from freshwater habitats. *Mycoscience* **40**: 165-183.
- Jones, E.B.G. 2011. Fifty years of marine mycology. *Fungal Divers.* **50**: 73-112.
- Jones, E.B.G. and K.L. Pang. 2010. Guest editorial: 11th International Marine and Freshwater Mycology Symposium, Taichung, Taiwan R.O.C., November 2009. *Bot. Mar.* **53**: 475-478.
- Jones, E.B.G. and S.T. Moss. 1978. Ascospore appendages of marine ascomycetes: an evaluation of appendages as taxonomic criteria. *Mar. Biol.* **49**: 11-26.
- Jones, E.B.G., J. Sakayaroj, S. Suetrong, S. Somrithipol, and K.L. Pang. 2009. Classification of marine Ascomycota, anamorphic taxa and Basidiomycota. *Fungal Divers.* **35**: 1-187.
- Jones, E.B.G., K.D. Hyde, S.J. Read, S.T. Moss, and S.A. Alias. 1996. *Tirisporella* gen. nov., an ascomycete from the mangrove palm *Nypa fruticans*. *Can. J. Bot.* **74**: 1487-1495.
- Jones, E.B.G., S.T. Moss, and V. Cuomo. 1983. Spore appendage development in the lignicolous marine pyrenomycetes *Chaetosphaeria chaetosa* and *Halosphaeria trullifera*. *Trans. Br. Mycol. Soc.* **80**: 193-200.
- Koch, J. and K.R.L. Petersen. 1996. A check list of higher marine fungi on wood from Danish coasts. *Mycotaxon* **15**: 397-414.
- Kohlmeyer, J. 1964. A new marine Ascomycete from wood. *Mycologia* **56**: 770-774.
- Kohlmeyer, J. and E. Kohlmeyer. 1979. *Marine Mycology: the Higher Fungi*. Academic Press, New York.
- Luo, W., L.L.P. Vrijmoed, and E.B.G. Jones. 2005. Screening of marine fungi for lignocellulose-degrading enzyme activities. *Bot. Mar.* **48**: 379-386.
- Pang, K.L., M.W.L. Chiang, and L.L.P. Vrijmoed. 2008. *Havispora longyearbyenensis* gen. et sp. nov.: an Arctic marine fungus from Svalbard, Norway. *Mycologia* **100**: 291-295.
- Pang, K.L., M.W.L. Chiang, and L.L.P. Vrijmoed. 2009. *Remispora spitsbergenensis* sp. nov., a marine lignicolous ascomycete from Svalbard, Norway. *Mycologia* **101**: 531-534.
- Pang, K.L., S.A. Alias, M.W.L. Chiang, L.L.P. Vrijmoed, and E.B.G. Jones. 2010. *Sedecimiella taiwanensis* gen. et sp. nov., a marine mangrove fungus with 16 spores in an ascus. *Bot. Mar.* **53**: 493-498.
- Pang, K.L., R.K.K. Chow, C.W. Chan, and L.L.P. Vrijmoed. 2011a. Diversity and physiology of marine lignicolous fungi in Arctic waters: a preliminary account. *Polar Res.* **30**: 1-5.
- Pang, K.L., J.S. Jheng, and E.B.G. Jones. 2011b. *Marine mangrove fungi in Taiwan*. National Taiwan Ocean University Press, Keelung, Taiwan, R.O.C.
- Shearer, C.A. and M. Miller. 1977. *Fungi of the Chesapeake Bay*

- and its tributaries V. *Aniptodera chesapeakensis* gen. et sp. nov. *Mycologia* **69**: 887-898.
- Spatafora, J.W., B. Volkmann-Kohlmeyer, and J. Kohlmeyer. 1998. Independent terrestrial origins of the Halosphaeriales (marine Ascomycota). *Am. J. Bot.* **85**: 1569-1580.
- Vrijmoed, L.L.P. 2000. Isolation and culture of higher filamentous fungi. In K.D. Hyde and S.B. Pointing (eds.), *Marine mycology-A practical approach*. Fungal Diversity Press, Hong Kong, pp. 1-20.

## 臺灣岩岸新種海洋海殼菌科子囊真菌 *Pileomyces formosanus* gen. et sp. nov.

彭家禮 鄭仁勝

國立臺灣海洋大學 海洋生物研究所

臺灣對於海洋真菌研究是不足的，最近的研究紀錄了紅樹林環境中 69 種海洋真菌，其中 34 種是新紀錄種。相對於紅樹林的研究，關於臺灣海洋岩岸海洋真菌多樣性之文獻是匱乏的。在新北市鶯歌石的一次野外採集調查中，發現了一種屬於海殼菌科 (Halosphaeriaceae) 的新種海洋真菌，其特徵為具黑色孢子器 (ascmata)，孢子囊 (asci) 為棒狀，子囊頂端較厚且具開口，內細胞質膜呈收縮狀 (retraction of plasmalemma)，孢子 (ascospores) 為橢圓形，其一端具有片狀附著器 (sheet-like appendage)，且會脫落。由於此新種的附著器型態在海殼菌科中是相當獨特的，因此建立一個新屬種 *Pileomyces formosanus*。 *Pileomyces* 屬之孢子囊形態和 *Aniptodera* 及 *Phaeonectriella* 兩屬相似，而孢子之附著器形態跟 *Tirisporella* (*T. beccariana*) 類似，皆存在於孢子一端。

**關鍵詞：**小囊菌目；海殼菌科；海洋真菌；漂流木；多樣性；分類。

