# STUDIES ON THE STREPTOMYCES SC<sub>4</sub> III. Biological Properties of Antibiotic SC<sub>4</sub>-X<sup>1,2</sup>

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#### Abstract

SC<sub>4</sub>-X, a novel antiliotic isolated from *Streptomyces* SC<sub>4</sub>, inhibits the growth of a variety of Gram-positive, Gram-negative bacteria and plant pathogenic fungi. The antimicrobial activity of SC<sub>4</sub>-X was influenced by pH of the test medium. SC<sub>4</sub>-X is more stable in neutral and acidic than in alkaline conditions, but it is more active in alkaline than in acidic solutions. SC<sub>4</sub>-X survived upon heating at 60°C for 10 hours and under UV irradiation for 24 hours. No degradation of SC<sub>4</sub>-X occurred by incubating with homogenates of rat heart, lung and kidney at 37°C for 120 minutes. No cross resistance between SC<sub>4</sub>-X, penicillin G and streptomycin was observed. The stepwise type of resistance development was similar to that of penicillin G. SC<sub>4</sub>-X was therapeutically effective against experimental *Staphylococcus aureus* avian infection in chickens.

Key words: Streptomyces; new streptothricin antibiotic; antibiotic activity; biological property.

# Introduction

The SC<sub>4</sub>-X is a new member of streptothricin antibiotics produced by a novel strain of *Streptomyces*, designated as SC<sub>4</sub>. Biological characters of this organism were reported by Wu (1984). Purification and chemical formulation of the SC<sub>4</sub>-X was also described (Wu *et al.*, 1983). In this paper, the biological activities and properties of SC<sub>4</sub>-X are reported.

### Materials and Methods

# Antibiotics

Antibiotic SC<sub>4</sub>-X was prepared as described previously by Wu *et al.* (1983). Antibiotic SC<sub>4</sub>-X was dissolved in phosphate buffer saline at pH 7 for *in vitro* and *in vivo* studies.

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The crystalline dihydrostreptomycin sulfate was purchased from Meiji Seika Kaisha, Japan. Penicillin G was purchased from Banyu Company, Japan.

# Determination of Minimum Inhibitory Concentration

Bacteria were cultivated on Difco nutrient agar, trypticase soy agar, antibiotic medium 1 or nutrient broth. The human pathogenic fungi were cultivated on Difco sabouraud medium. The Czapek-Dox medium was used for culturing plant pathogenic fungi and true fungi. Antibiotic  $SC_4$ -X and other test agents were dissolved in sterile water for serial two fold dilution with initial concentration at 1,000 mcg/ml. Antibacterial and antifungal activities of the test antibiotics were assayed by the paper disc method. Aliquots of  $5\,\mu$ l solutions were applied on the paper discs, 6 mm in diameter, which were then placed on soft agar seeded with appropriate test organisms. Non-pathogenic bacteria were incubated at 30°C and human pathogenic bacteria were incubated at 37°C for 18-24 h. True fungi were incubated at 28°C for 4 to 5 days. The minimum inhibitory concentration (MIC) was determined as the lowest concentration at which the visible growth of the test organism was completely inhibited. The diameter of each inhibition zone was measured by a caliper.

# Influence of Various Conditions on Antimicrobial Activities of SC<sub>4</sub>-X

Effect of pH on activity: The MIC of SC<sub>4</sub>-X against bacteria at different pH was determined by the paper disc method. The pH of hard agar medium and soft agar medium was adjusted to pH 6,7, and 8 with HCl or NaOH before sterilization. Six different microorganisms, Escherichia coli NIHJ, Bacillus subtilis ATCC 6633, Sarcina lutea ATCC 9341, Shigella flexneri 3B, Mycobacterium pseudotuberculosis 607, and Penicillium digitatum were used as indicator organisms.

Effect of inoculum size: The MIC of SC<sub>4</sub>-X against bacteria at different inoculum size was determined by the paper disc method on nutrient agar with inoculum sizes of 10<sup>2</sup>, 10<sup>4</sup>, 10<sup>6</sup>, and 10<sup>7</sup> viable cells per ml.

Effect of serum: The effect of animal serum on the MIC of SC<sub>4</sub>-X against bacteria was determined by paper disc method on trypticase soy agar plates containing 0%, 10%, 20% and 50% by volume of pig serum seeded with 10<sup>6</sup> viable cells per ml of Staphylococcus aureus avian or Escherichia coli NIHJ.

# Stabilitly of SC<sub>4</sub>-X in Various Solution at 37°C

SC<sub>4</sub>-X was dissolved in phosphate buffer saline at pH 7, trypticase soy broth, human blood or human urine to give concentration of SC<sub>4</sub>-X at 100 mcg/ml. Solutions were kept at 37°C for 5 days. The residual activity of SC<sub>4</sub>-X in the solution was assayed daily by paper disc method.

# Stability of SC<sub>4</sub>-X in Phosphate Buffer at Various pH

Antibiotic SC<sub>4</sub>-X was dissolved in phosphate buffer saline at pH 5, 7 and 9 to give concentration of 100 mcg/ml. Solutions were kept at 60°C for 52 h. SC<sub>4</sub>-X activity was assayed by paper disc method every 4 h.

#### Thermostability Test

Antibiotic SC<sub>4</sub>-X was dissolved in phosphate buffere saline at pH 7 to 100 mcg/ml and maintained at 60°C, 37°C and 10°C for 48 h in a water bath. At various time intervals till 48 h, 0.1 ml of each sample was withdrawn and assayed for antimicrobial activity by the paper disc method.

#### UV Stability Test

Antibiotic  $SC_4$ -X was dissolved in phosphate buffered saline at pH7 to a final concentration of 100 mcg/ml and the solution was irradiated by a UV lamp at a distance of 10 cm with constant stirring.  $SC_4$ -X residual activity was measured every 60 minutes by paper disc method. Ultraviolet light was emitted from a mercury vapor quartz tube that centered at 2,537 Å with intensity of about 155  $\mu$  watts/cm² at 44.5 cm distance.

# Stability of SC<sub>4</sub>-X in Tissue Homogenates

To estimate the stability of  $SC_4$ -X in tissue homogenates, male Wister rat about 200 g body weight were used. Twenty percent tissue homogenates each of liver, kidney, lung, heart and small intestine were prepared in 67 mM phosphate buffered saline at pH7. One ml of 1,000 mcg/ml of  $SC_4$ -X antibiotic solution was mixed with 9 ml of the above tissue homogenates, respectively. The mixture was incubated at 37°C for 30, 60, 90, or 120 minutes. Incubation was stopped by adding 2 ml of 99% ethanol to 1 ml of above reaction mixture. The residual activity of  $SC_4$ -X in the mixture was determined by the paper disc method.

# Development of Resistance

Patterns of development of resistance in *Staphylococcus aureus* 209 p against  $SC_4$ -X, streptomycin and penicillin G was studied using Difco trypticase soy broth and trypticase soy agar plates. *Staphylococcus aureus* 209p was incubated in trypticase soy broth overnight and 0.1 ml of culture suspension at  $10^7$  vc/ml was plated on trypticase soy agar plates containing  $1, 2, 4, \ldots, 1,000$  mcg per ml of antibiotics. After 48 h of incubation at  $37^{\circ}$ C, ten colonies on plates with the highest concentration of antibiotic were isolated and inoculated in trypticase soy broth at  $37^{\circ}$ C for 24 h. The suspensions were reinoculated on the next series of plates. Transfers were made every 48 h from the tube containing the highest concentration of

the antibiotic permitting growth into the next series of plates containing the same and several higher concentrations of antibiotics.

# Bactericidal Activity of SC<sub>4</sub>-X

The viability of Staphylococcus aureus 209p in the presence of SC<sub>4</sub>-X was determined by the plate count technique. A 24 h broth culture of Staphylococcus aureus 209p was diluted 1,000 times in trypticase soy broth and the antibiotic was added to give concentrations of 1,000, 100, 10, 1.0, 0.1, and 0.01 mcg per ml. One ml of culture broth was withdrawn at intervals of 2, 4, 6 and 8 h from each tube prior to incubation at 37°C. Cells were washed with 10 ml of sterile normal saline three times to remove residual antibiotic from cells. Cells prepared as above were inoculated on trypticase soy agar plates. Colony counts were made after 48 h of incubation.

#### Cross Resistance

Cross resistance between SC<sub>4</sub>-X and other antibiotics was studied with Staphylococcus aureus 209p. The bacteria were made resistant to SC<sub>4</sub>-X, streptomycin and penicillin G respectively by spontaneous mutation and selection method. These mutants were cultivated on trypticase soy agar plates respectively. The MIC of SC<sub>4</sub>-X, penicillin and streptomycin on the cell were then determined by paper disc methods.

#### Acute Toxicity in Mice

White male ICR mice, 20 grams weight, were used. Animals were divided into 10 groups, each with 10 mice, and injected subcutaneously with various doses of SC<sub>4</sub>-X antibiotic. The 50 percent lethal dose (LD<sub>50</sub>) was calculated from the survival rate of the mice within 7 days of injection.

#### Experimental Infection of Chieken

Staphylococcus aureus (avian type) was cultivated on trypticase soy agar slants for 20 h. Cells were then harvested and washed 3 times with saline and finally suspended in normal saline to make cell suspensions of  $5\times10^7$  per ml. Each 0.2 ml cell suspension was injected subcutaneously into chickens. At the same time, each inoculated chicken was administered subcutaneously with various dose of  $SC_4$ -X antibiotic. The symptoms were observed by eyes.

#### Results

# Antibacterial Spectrum

The antimicrobial spectrum and minimum inhibitory concentration of SC<sub>4</sub>-X against bacteria, animal pathogenic bacteria and fungi are summarized in Table 1.

Table 1. Minimal inhibitory concentrations of antibiotic SC<sub>4</sub>:X

Test organism	Medium	MIC in mcg/ml	Incubation time	
Staphylococcus aureus ATCC 65389	Antibiotic medium 1	15.1	24 h at 37°C	
Sarcina lutea ATCC 9341	Antibiotic medium 1	1.4	24 h at 37°C	
Bacillus subtilis PCI 219	Antibiotic medium 1	15.6	24 h at 37°C	
Bacillus subtilis ATCC 6633	Antibiotic medium 1	3.9	24 h at 37°C	
Corynebacterium xeloses	Antibiotic medium 1	3.9	24 h at 37°C	
Shigella flexneri 3B	Antibiotic medium 1	3.9	24 h at 37°C	
Escherichia coli NIHJ	Antibiotic medium 1	1.95	24 h at 37°C	
Proteus vulgaris	Antibiotic medium 1	100	24 h at 37°C	
Pseudomonas aeruginosa	Antibiotic medium 1	100	24 h at 37°C	
Klebsiella pneumoniae ATCC 10031	Antibiotic medium 1	100	24 h at 37°C	
Mycobacterium pseudotuberculosis 607	Antibiotic medium 1 with 5% glycerin	3.9	5 days at 37°C	
Escherichia coli swine	Trypticase soy agar $+10\%$ pig serum and $+10\%$ yeast extract	15.6	24 h at 37°C	
Staphylococcus aureus avian	Trypticase soy agar +10% pig serum and +10% yeast extract	7.8	24 h at 37°C	
Bordetella bronchiseptica	Trypticase soy agar +10% pig serum and +10% yeast extract	31.2	24 h at 37°C	
Hemophilus pleuropneumonia	Trypticase soy agar +10% pig serum and +10% yeast extract	7.8	24 h at 37°C	
Pasteurella hemolytica	Trypticase soy agar +10% pig serum and +10% yeast extract	62.5	24 h at 37°C	
Aeromonas hydrophila	Trypticase soy agar +10% NaCl	62.5	24 h at 28°C	
Vibrio anguillarum	Trypticase soy agar +10% NaCl	100	24 h at 28°C	
Saccharomyces cerevisiae	Sabouraud agar	7.8	5 days at 30°C	
Cryptococcus neoformans	Sabouraud agar	32	5 days at 37°C	
Candida albicans	Sabouraud agar	32	5 days at 37°C	
Aspergillus niger	Czapek-Dox agar	100	5 days at 28°C	
Colletotricum musae	Czapek-Dox agar	100	5 days at 28°C	
Colletotricum lageunarium	Czapek-Dox agar	31.2	5 days at 28°C	
Fusarium moniliforus	Crapek-Dox agar	62.5	5 days at 28°C	
Gibberella fujikuroi	Czapek-Dox agar	15.6	5 days at 28°C	
Penicillium citrinum	Czapek-Dox agar	15.6	5 days at 28°C	
Penicillium digitatum	Czapek-Dox agar	15.6	5 days at 28°C	
Penicillium italicum wehmer	Czapek-Dox agar	31.2	5 days at 28°C	
G'oeosporium papayae	Czapek-Dox agar	31.2	5 days at 28°C	

Inoculum size: 10<sup>6</sup> vc/ml. Method: paper disc method. Antibiotic SC<sub>4</sub>-X showed strong antimicrobial activity against several bacteria, such as Sarcina lutea, Corynebacterium xelosis, Escherichia coli NIHJ, Mycobacterium pseudotuberculosis 607, Staphylococcus aureus avian and some fungi such as Saccharomyces cerevisiae, Gibberella fujikuroi and Penicillium citrinum etc. On the other hand, SC<sub>4</sub>-X showed poor antimicrobial activity against Gram-negative bacilli, such as Proteus vulgaris, Pseudomonas aeruginosa and Klebsiella pneumoniae.

# Influence of Culture Conditions on Antibacterial Activity

The MIC of SC<sub>4</sub>-X against various organisms under various conditions were observed.

Inoculum size: The activity of  $SC_4$ -X was obviously influenced by the inoculum size. Table 2 indicated clearly that the antibacterial activity increased as the inoculum size decreased.

Table 2. Effect of inoculum size on antibacterial activity of SC<sub>4</sub>-X

Organism	MIC in mcg/ml				
	Inoculum size vc/ml				
	107	10 <sup>6</sup>	104	10 <sup>2</sup>	
Salmonella paratyphi B.	7.75	3.9	1.95	1.95	
Sarcina lutea ATCC 9341	31.25	15.5	1.95	1.98	
E. coli NIHJ	15	7.75	7.75	1.95	
Bacillus subtilis PIC 219	7.75	3.9	0.95	0.95	

Medium: trypticase soy agar (Difco) at pH 7.

Method: paper disc method. Incubation: at 37°C for 18 to 24 h.

pH effect: The results are shown in Table 3. In most of the organisms tested, the MIC of  $SC_4$ -X antibiotic at pH6 were about ten times higer than that at pH8.

Binding with serum protein: Effect of the addition of serum to the activity of SC<sub>4</sub>-X is shown in Table 4. Antimicrobial activity against Staphylococcus aureus avian and Escherichia coli NIHJ was the same as the medium in the presence of pig serum up to 10%. This shows that the antibiotic SC<sub>4</sub>-X does not severely bind with serum proteins.

# Stability of SC<sub>4</sub>-X Under Various Experimental Conditions

Figure 1a shows the residual activity of  $SC_4$ -X in various aqueous conditions at 37°C.  $SC_4$ -X was very stable in phosphated buffer at pH 7.0 and 37°C up to 5 days. About 50%  $SC_4$ -X activity in trypticase soy broth and 95% in human urine were inactivated after incubating at 37°C for 4 days.

Table 3. Effect of medium pH on antimicrobial activity of SC<sub>4</sub>X

	Organism	Medium	M.I.C. in mcg/ml		g/ml
	Organism	Medium	pH 6	pH 7	рН 8
٠.,	Escherichia coli NIHJ	A	31.2	7.8	2.0
	Bacillus subtilis ATCC 6633	A	31.2	15.6	3.9
	Sarcina lutea ATCC 9341	A	15.6	3.9	2.0
	Shigella flexneri 3B	A	31.2	7.8	3.9
	Mycobacterium pseudotuberculosis 607	NG	31.2	1	1
	Penicillum digitatum	С	125	15.6	15.6

A: Difco antibiotic medium 1.

NG: Difco nutrient agar with 5% of glycerol.

Inoculum size:  $5 \times 10^6$  vc/ml.

Paper disc method: Common bacteria were incubated at 37°C for 24 h. Mycobacterium was incubated at 37°C for 5 days. Penicillum was incubated at 28°C for 5 days.

**Table 4.** Effect of pig serum concentration in medium on antibacterial activity of  $SC_4$ -X

Organism	Serum (%)	MIC in mcg/m
Staphylococcus aureus avian	0	7.75
	. 1	7.75
	10	7.75
	20	7.75
	50	7.75
Escherichia coli NIHJ	0	7.75
	1	7.75
	10	7.75
	20	7.75
	50	7.75

Inoculum size: 10<sup>6</sup> vc/ml. Method: paper disc method. Medium: nutrient agar.

Staphylococcus aureus (avian type) was isolated from poultry farm.

The residual activity of  $SC_4$ -X in phosphate buffer of different pH at  $60^{\circ}C$  was shown in Fig. 1b. Antibiotic  $SC_4$ -X was more stable in neutral and acidic than in alkaline conditions. The activity of  $SC_4$ -X in solution remained constant at pH 5 and 7 up to 12 h but diminished rapidly at pH 9.

The stability of SC<sub>4</sub>X at different temperatures is shown in Fig. 1c. SC<sub>4</sub>X

C: Czapek-Dox agar.

maintained high stability at 10°C and only 25% of activity was loss after heating at 60°C for 10 h.

As shown in Fig. 1d,  $SC_4$ -X was stable under UV irradiation. It indicates that the  $SC_4$ -X is insensitive to UV light.

As shown in Fig. 1e, no degradation of SC<sub>4</sub>-X was occurred by incubating it with animal heart, lung and kidney homogenates for 120 minutes, while about 25% of SC<sub>4</sub>-X activity was inactivated in intestinal homogenate.

#### Cross Resistance

Cross resistance between SC<sub>4</sub>-X, penicillin G and streptomycin was studied on Staphylococcus aureus 209p which was made resistant to Sc<sub>4</sub>-X, penicillin G or streptomycin respectively by serial subculture in trypticase soy agar plates containing various concentrations of each antibiotic. Table 5 shows that SC<sub>4</sub>-X maintoined similar activity against microorganisms, which was made resistant to other antibiotics and SC<sub>4</sub>-X resistant organism was still sensitive to other antibiotics. It indicates that no cross resistant occurred among penicillin G, streptomycin and SC<sub>4</sub>-X antibiotic.

**Table 5.** Cross resistance test among SC<sub>4</sub>·X, pencillin G and dihydrostreptomycin

	MIC			
Organism —	SC <sub>4</sub> -X	Pe.	Strep.	
Sta. aureus 209 P parent type	15.6	0.1	20	
R-SC <sub>4</sub> -X	30	0.1	20	
R-penicillin G	15.6	31	20	
R-streotomycin	15.6	0.1	500	

Inoculum size: 106 vc/ml.

Method: paper disc method.

Medium: trypticase soy agar (Difco).

### Development of Resistance

The development of resistance of *Staphylococcus aureus* 209p to SC<sub>4</sub>-X, penicillin G and streptomycin was compared. The progression and degree of resistance to 3 kinds of antibiotics are shown in Fig. 2. For streptomycin resistance, the MIC of streptomycin increased from 50 mcg/ml to 500 mcg/ml needs only one transfer and was not affected by further transfer. Patterns of gradual development of resistance against SC<sub>4</sub>-X and penicillin G are similar. But resistant against penicillin G was more rapidly developed than those against SC<sub>4</sub>-X.

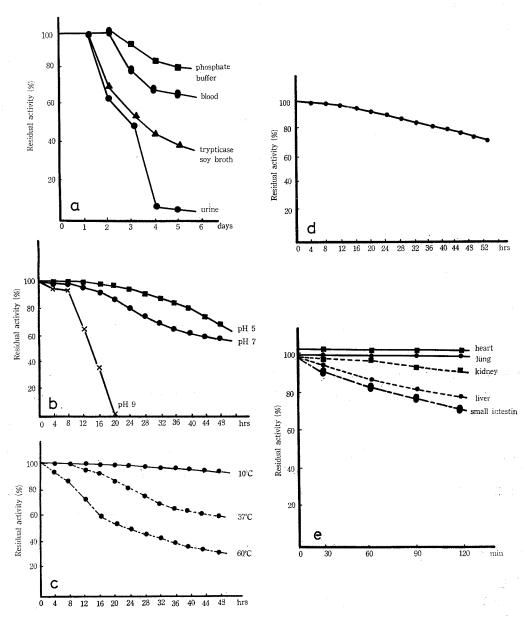


Fig. 1. Stability of SC<sub>4</sub>-X under various experimental conditions.

- (1a) Residual activity of SC<sub>4</sub>-X in various aqueous conditions at 37°C.
- (1b) Residual activity of SC<sub>4</sub>-X in phosphate buffer of different pH at 60°C.
- (1c) Residual activity of SC<sub>4</sub>-X at different temperatures. The antibiotic SC<sub>4</sub>-X was dissolved in phophate buffer at pH. 7.0.
- (1d) Sensitivity of SC<sub>4</sub>-X to UV illumination.
- (1e) Residual activity of  $SC_4$ -X in animal tissue homogenates.

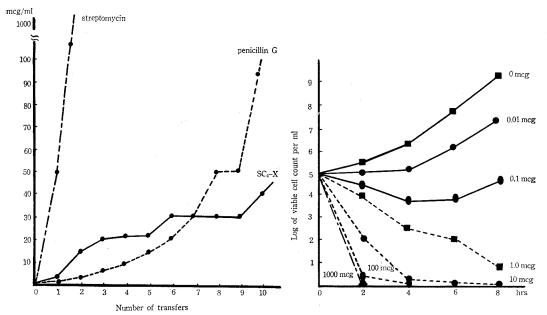


Fig. 2. Patterns of development of resistance of Staphylococcus aureus 209 P to SC<sub>4</sub>-X, stretomycin and penicillin G.

Fig. 3. Bactericidal effects of SC<sub>4</sub>-X against Staphylococcus aureus 209 P.

#### Bactericidal Activity

The viability of *Staphylococcus aureus* 209p cultivated in trypticase soy broth containing various concentrations of SC<sub>4</sub>-X was determined by plate count, as illustrated in Fig. 3. The logarithm of viable cell counts were plotted against exposure time to antibiotic. When the antibiotic was added simultaneously, weak bacteriocidal action was observed at concentration of 10 mcg/ml. At concentration of 1 mcg/ml, the viable cell counts did not change through a period of 8 h. The increase of viable cell count at the concentration of 0.01 mcg/ml of SC<sub>4</sub>-X was only slightly less than that in control. These results reveal that SC<sub>4</sub>-X is rather bacteriostatic than bacteriocidal.

### LD50 Value

At a dosage of 250 mcg/g body weight,  $SC_4$ -X was not lethal to mice, but animals were severely ill even on the 5th day after injection. All mice received 300 mcg/g body weight died within 5 days. So, the  $LD_{50}$  value for  $SC_4$ -X to mice ICR was 275 mcg/g body weight.

# Experimental Infection of Chicken

Antibiotic SC<sub>4</sub>-X was simultaneously administered to chicken and challenged intramuscularly with the strain of Staphylococcus aureus avian type. Positive effect

was obtained with a dose of 10 mcg/g body weight of  $SC_4$ -X in the chicken, exhibiting significantly preventive effect.  $SC_4$ -X was not harmful to chicken at concentration of 50 mcg/g body weight.

#### Discussion

Since streptothricin F was discovered by Waksman and Woodruff in 1942, investigation of streptothricins have been processed rather slowly. After prolonged investigations by American, British and Japanese scientists, the structures of five first ninhydrin-positive compounds isolated from the total and partial hydralysates of streptothricins were established by Khokhlov et al. in 1972. Accordingly, all streptothricins and streptothricin-like antibiotics are active against a wide spectrum of Gram-positive, Gram-negative bacteria and fungi, but are highly nephrotoxic to animals (Khokhlov et al., 1961), which limited their practical use in medicine. Their application in agriculture seems more promising both for suppression of some phytopathogenic microorganisms and as supplement into animal feeds.

In our continuous search for antibiotics of anti-fungi and animal feed additive,  $SC_4$ -X was isolated from *Streptomyces*  $SC_4$  as a new member of water-soluble streptothricin-type antibiotics. *In vitro* tests show that  $SC_4$ -X is active against common bacteria, phytopathogenic fungi and animal pathogenic bacteria.

Although many antibiotics are available for the treatment of infectious disease in livestock and poultry, prolonged treatment causes resistance. We are particularly aware of infection of Staphylococcus in chicken in Taiwan, and its gradual development of resistance that causes penicillin less effective. SC4-X is effective against Staphylococcus aureus in chicken and have the potiential to treat Staphylococcus aureus infection in poultry. SC4-X is reasonably stable at elevated temperature, under UV irradiation and in incubation with animal tissue homogenates. Although SC4-X is more stable under acidic conditions it is more active as its free base form in alkaline solution. This trend of stability is advantageous for SC4-X in its practical application. Animals normally can maintain quite well under neutral or slightly alkaline physiological conditions to give SC<sub>4</sub>-X long acting period. SC4-X is less likely to develop resistance than streptomycin from Staphylococcus aureus 209p. The stepwise type of resistance development in Staphylococcus against SC<sub>4</sub>-X is characteristic and no cross resistance between SC<sub>4</sub>-X, penicillin G and streptomycin were observed in our studies. From these points of view, SC4-X has great potential to develop as antibiotic for animal use.

#### Literature Cited

Akasak, K., H. Abe, A. Seino, and S. Shiratol. 1968. Yazumycin, a new antibiotic produced by *Streptomyces lavendulae*. J. Antibiotics 21: 98-105.

- Baker, H., A. Sidorowicz, S. N. Schgal, and C. Vezina. 1978. Rapamycin (AY-22, 989), a new antifungal antibiotic. III. *In vitro* and *in vivo* evaluation. J. Antibiotics 31: 539-545.
- Brown, W.E., J. Szanto, E. Meyers, T. Kawamura, and K. Arima. 1977. Taeniacidal activity of Streptothricin antibiotic complex S15-1 (SQ 21, 704). J. Antibiotics 30: 886-889.
- Carter, E., R.K. Clark, P. Koine, J.W. Rothrock, W.R. Tayler, C.A. West, G.B. Whitfield, and G. Jackson. 1954. Strptothricin I. Preparation, properties and hydrolysis products. J. Am. Chem. Soc. 76: 566-569.
- Carter, H. H., C. C. Sweeley, E. E. Daniels, J. E. Mcnary, C. P. Schaffner, C. A. West, E. E. Tamelen, J. R. Dyer, and H. A. Whaley. 1961. Streptothricin and streptolin: The structure of streptolidine (Roseonine). J. Am. Chem. Soc. 83: 4296-4297.
- Fried, J. and O. Wintersteiner. 1945. Crystalline reineckates of streptothricin and streptomycin. Science 101: 613-615.
- Goto, T. and T. Ohgi. 1974. Synthesis of roseonine (streptolidine), a guanidino-amino acid component of streptothricin group antibiotics. Tetrahedran Lett. 15: 1413-1416.
- Goto, S. and S. Kuwahara. 1968. *In vitro* and *in vivo* evaluation of enduracidin, a new peptide antibiotic substance. J. Antibiotics 21: 119-125.
- Haneisih, H., M. Arai, N. Kitano, and S. Yamamoto. 1974. Aspiculamycin, a new cytosine nucleoside antibiotic. III. Biological activities, in vitro and in vivo. J. Antibiotics 27: 339-342.
- Hamilton-Miller, J. M. T., D. W. Kerry, and W. Brumfitt. 1974. An *in vitro* comparison of cefoxitin, a semi-synthetic cephamycin, with cephalothin. J. Antibiotics 27: 42-48.
- Ito, Y., Y. Ohashi, Y. Sakurai, M. Sakurazawa, H. Yoshida, S. Awataguchi, and T. Okuda. 1968. New basic water-soluble antibiotics BD-12 and By-81 II Isolation, purification and properties. J. Antibiotics 21: 307-313.
- Iwata, K., Y. Yamamoto, H. Yamaguchi, and T. Hiratani. 1982. In vitro studies of aculeacin A, a new antifungal antibiotic. J. Antibiotics 35: 203-218.
- Kawakami, M., Y. Nagai, T. Fujii, and S. Mitsuhasii. 1971. Anti-microbial activity of enduracin (Enramycin) in vitro and in vivo. J. Antibiotics 34: 583-586.
- Kawaharajo, K., Y. Sekizawa, and M. Inoue. 1081. In vitro and in vivo antibacterial activity of 9, 3"-di-O-acetyl medecamycin (MOM), A new macrolide antibiotic. J. Antibiotics 34: 436-442.
- Kondo, F., N. Kitano, and H. Domon. 1974. Aspiculamycin, a new cytosine neucleside antibiotic IV. Antimycoplasma activity of aspiculamycin in vitro and in vivo. J. Antibiotics 27: 529-534
- Kondo, M., T. Oishi, K. Ishifuji, and K. Tsuchiya. 1973. Maridomycin, a new macrolide antibiotic III. *In vitro* and *in vivo* antibacterial activity. J. Antibiotics 36: 206-214.
- Johnson, A. W., and J. W. Westley. 1962. Streptothricin group of antibiotics. Part 1. The general structural pattern. J. Chem. Soc. 1642-1652.
- Kawamura, T., K. Tago, T. Beppu, and K. Arima. 1976. Antiviral antibiotic S15-1. J. Anti-biotics 29: 242-247.
- Kawamura, T., T. Kimura, T. Tago, T. Beppu, and K. Arima. 1976. The identity of S15-1-A and B with racemomycins A and C. J. Antibiotics 29: 845-846.
- Khokhlov, A.S., and P.D. Reshetov. 1964. Chromatography of streptothricins of carboxymethylcellulose. J. Chromatography 14: 495-496.
- Khokhlov, A.S., and K.I. Shutova. 1972. Chemical structure of structure of streptothricins. J. Antibiotics 25: 501-508.
- Khokhlov, A.S. 1961. Chemistry of antibiotics. Publ. House of USSR, Acade. Sci. Moscow.
- Komatsu, N., Y. Saburi, Y. Hirata, T. Mizuno, and N. Sakai. 1952. Counter-current distribution studies of H-277. J. Antibiotics 9: 522-523.
- Kono, Y., S. Makino, S. Takenchi, and H. Yonehara. 1969. Sclerothricin, a new basic antibiotic. J. Antibiotics 22: 583-589.
- Larson, L. M., H. Sternberg, and W. H. Peterson. 1953. Production, isolation and components of the antibiotic streptolin. J. Am. Chem. Soc. 75: 2036-2039.

- Mine, Y., T. Kamimura, S. Nonoyama, and M. Nishida. 1980. In vitro and in vivo antibacterial activities of FR-31564, a new phosphonic acid antibiotic. J. Antibiotics 33: 36-43.
- Nakanishi, K., T. Ito, and Y. Hirata. 1954. Structure of a new amino acid obtained from roseothricin. J. Am. Chem. Soc. 76: 2845-2846.
- Noto, T., T. Nehashi, H. Endo, M. Saito, S. Matsubara, Y. Harada, S. Suzuki, H. Ogawa, and K. Koyama. 1976. Ceftezole, a new cephalosporin C derivative I. In vitro and in vivo antimicrobial activity. J. Antibiotics 24: 1058-1066.
- Peck, R. L., A. Walti, R. P. Graber, E. Flynn, C. E. Hoffhine, V. Allfrey, and K. Folkers. 1946. Streptomyces antibiotics VII. The structure of streptidine. J. Am. Chem. Soc. 68: 776-781.
- Rivett, R. W. and D. H. Peterson. 1947. Streptolin, a new antibiotic from a species of *Streptomyces*. J. Am. Chem. Soc. 69: 3006-2009.
- Sawada, Y., H. Taniyama, N. Hanyuda, H. Hayashi, and T. Ishida. 1974. A new streptothricin antibiotic R 4H. J. Antibiotics 27: 535-543.
- Sawada, Y., H. Sakamoto, and H. Taniyama. 1974. Studies on chemical modification of streptothricin-group antibiotics III. Partial N-acetylation of racemomycin and their biological activity. J. Pharma. Soc. Japan 94: 176-180.
- Sawada, Y. and H. Taniyama. 1974. Studies on chemical modification of streptothricin-group antibiotics. IV. Preparation of  $\beta$ -N-Acetylracemomycin-A derivative and its antimicrobial activity. J. of Pharma. Soc. Japan 94: 264-266.
- Sawada, Y. and H. Taniyama. 1974. Studies on chemical modification of streptothricin-group antibiotics. V. Synthesis of amino acid derivative on β-lysine of racemomycin A and their biological activity. J. Pharm. Soc. Japan 94: 858-864.
- Sawada, Y., S. Kawakami, and H. Taniyama. 1977. Glycinothricin, a new streptothricin-class antibiotics from *Streptomyces griscus*. J. Antibiotics 30: 460-467.
- Shimazu, A., T. Hidaka, S. Otsuka, M. Nishiyama, and H. Yonehara. 1969. Streptomyces sclerogranulatus sp. Nov., The producer of sclerothricin. J. Antibiotics 22: 590-596.
- Shoji, S., S. Kozuki, M. Ebata, and H. Otsuka. 1968. A water-soluble basic antibiotic E-749-C identical with LL-AC 541. J. Antibiotics 21: 509-511.
- Tamelen, E.E. and E.E. Smissman. 1952. Streptolin, the structure and synthesis of isolysine. J. Am. Chem. Soc. 74: 3713-3714.
- Tamelen, E.E. and E.E. Smissman. 1953. Streptolin, the structure and synthesis of isolysine I. Am. Chem. Soc. 75: 2031-2035.
- Tamelen, E. E., J. R. Dyer, H. E. Carter, J. V. Pierce, and E. E. Daniels. 1956. Structure of the aminosugar derived from streptothricin and streptolin B. J. Am. Chem. Soc. 78: 4817-4818.
- Thiele, E. H. 1974. An *in vivo* pyelonephritis assay for screening therapeutic agents. J. Anti-biotics 27: 31-41.
- Van Tamelen, E. E., S. R. Dyer, H. A. Whaley, H. E. Carter, and G. B. Whitfield. 1961. Constitution of the streptolin-streptothricin group of *Streptomyces* antibiotics. J. Am. Chem. Soc. 83: 4295-4296.
- Taniyama, H. and S. Takemura. 1957. Chemical studies on antibiotics produced by *Actinomycetes*. I. Racemomycin (I), Isolation and purification of racemomycin B (229-B). J. Pharm. Soc. Japan 77: 1210-1214.
- Taniyama, H. and S. Takemura. 1957. Chemical studies on antibiotics produced by Actinomycetes. II. Racemomycin. (2). Hydrolysis of racemomycin B. (i). J. Pharm. Soc. Japan 77: 1215-1217.
- Taniyama, H. and S. Takemura. 1958. Chemical studies on antibiotics produced by Actinomycetes. III. Racemomycin (3). Hydrolysis of racemomycin B (ii). J. Pharm. Soc. Japan 78: 742-744.
- Taniyama, H., Y. Sawada, and T. Kitagawa. 1971. Chromatography of racemomycin on dextran gel. J. Chromatogr 56: 360-362.
- Taniyama, H., Y. Sawada, and T. Kitagawa. 1971. Studies on the inactivation and regeneration of streptothricin. J. Antibiootics 24: 662-666.
- Taniyama, H., Y. Sawada, and T. Kitagawa. 1971. The identity of yazumycins A and C with racemomycins A and C. J. Antibiotics 24: 390-392.

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- Taniyama, H., Y. Sawada, and K. Hasimoto. 1972. Studies on chemical modification of Streptothricin-group antibiotics. I. On citromycin derivatives. J. Pharm. Soc. Japan 92: 182-186.
- Tsuchiya, K., T. Yamazaki, Y. Takeuchi, and T. Oishi. 1971. Studies on T-2636 antibiotics. IV. In vitro and in vivo antibacterial activity of T-2636 antibiotics. J. Antibiotics 24: 29-41.
- Tsuchiya, K., M. Kondo, T. Oishi, and T. Yamazaki. 1968. Enduracidin, a new antibiotic III. *In vitro* and *in vivo* antimicrobial activity. J. Antibiotics 21: 147-153.
- Tsuruoka, T., T. Shoumura, N. Ezaki, T. Niwa, and T. Nhda. 1968. SF-701, a new strepto-thricin-like antibiotic. J. Antibiotics 21: 237-238.
- Vander Brook, M. J., A. N. Wick, W. H. Devries, R. Harris, and G. F. Cartland. 1946. Extracion and purification of streptomycin, with a note on streptothricin. J. Biol. Chemi. 165: 463-468.
- Waksman, S. A., and H.B. Woodruff. 1942. Streptothricin a new selective bacteriostatic and bactericidal agent, particularly active against gram-negative bacteria. Proc. Soci. Experi. Biol. and Medicin 49: 207-210.
- Waksman, S. A. 1943. Production and activity of streptothricin. J. Bacteriol. 46: 299-310.
- Wu, R.Y., M.C. Shiao, and H.M. Lee. 1983. Studies on the *streptomyces* SC<sub>4</sub>: Chemical formulation of antibiotic SC<sub>4</sub>-X. Bot. Bull. Academia Sinica 24: 71-87.
- Wu, R.Y. 1984. Studies on the streptomyces SC<sub>4</sub>. II. Taxonomic and biological characteristics of streptomyces strain SC<sub>4</sub>. Bot. Bull. Academia Sinica. 25: 111-123.
- Yamazaki, H. 1968. Studies on antimicrobial substance B 44P (streptovaricin) produced by a strain of Actinomycetes. III. Chemotherapeutic effect on Staphylococcal infection in mice. J. Antibiotics 21: 222-226.

# 鏈 黴 菌 SC<sub>4</sub> 之 研 究 III. SC<sub>4</sub>-X 抗生素之生物特性

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鏈黴菌  $SC_4$  是由臺灣土壤中分離而得之新抗生素產生菌。 $SC_4$ -X 抗生素可將  $SC_4$  菌培養液,經離子交換及層柱分析法予以純化。  $SC_4$ -X 抗生素抗菌譜甚廣,對革蘭氏陽性菌,陰性菌及植物病原性眞菌等具有生長抑制的能力。  $SC_4$ -X 之活性受培養基之酸鹼度所影響,其活性在鹼性下較酸性中爲强,但安定性則反之,在酸性下之抗生素較在鹼性中爲安定。 高溫處理及紫外光之照射均不影響  $SC_4$ -X 抗生素之活性。  $SC_4$ -X 不受白老鼠之心臟,肝臟及腎臟磨碎液破壞。  $SC_4$ -X 與盤尼西林 G 及鍵黴素之間無交叉抗藥性。葡萄球菌對  $SC_4$ -X 之抗藥性與盤尼西林類似,須經數次之接移才能產生。  $SC_4$ -X 抗生素對鶏葡萄球菌的感染預防及治療有良好的藥效。